

# mAb Glycopeptide Profiling with V-Tag

Adding reliable glycoprofiling  
to your peptide mapping workflow  
with ease and simplicity



# Who is the V-Tag Glycoprofiling Technology for?

V = Velocity

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**The Ludger V-Tag Glycoprofiling Technology is for mAb developers, both innovator and biosimilar companies, who need to glycoprofile\* mAb samples reliably, at an affordable cost and with a fast turnaround time.**

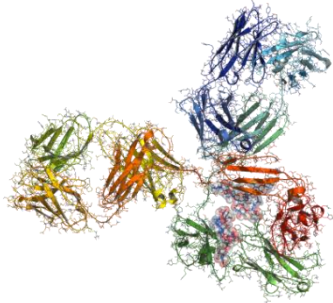
**Our typical V-Tag clients are those who want to:**

<b>Monitor their mAb drug's glycosylation during the product lifecycle. This includes showing the comparability of glycosylation throughout the drug lifecycle as well as biosimilarity to an innovator's drug</b>	<b>Integrate glycoprofiling into the peptide mapping workflow. The V-Tag labeled glycopeptides are analysed using orthogonal platforms, MALDI-MS and UHPLC</b>
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\*Glycoprofile = a map of the drug's glycosylation containing structural ID and relative abundance of each glycan species

# Options for Analysing mAb Glycosylation

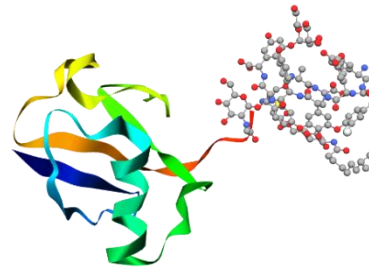
Three types of mAb-derived molecules can be analysed to gain information about mAb glycosylation



## 1. Intact Glycoprotein

*Using lectin affinity, CE or MS*

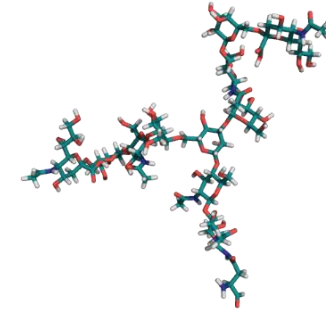
- ✓ Provides information on **molecular weight and glycoform distribution**
- X *Poor structural information for the glycans. Insufficient for regulatory work or lot release*



## 2. Glycopeptides

*Using MS and UHPLC*

- ✓ Provides both **glycan identity and quantitation** as well as **glycan attachment site and site occupancy**
- V-Tag is designed for glycopeptide mapping*



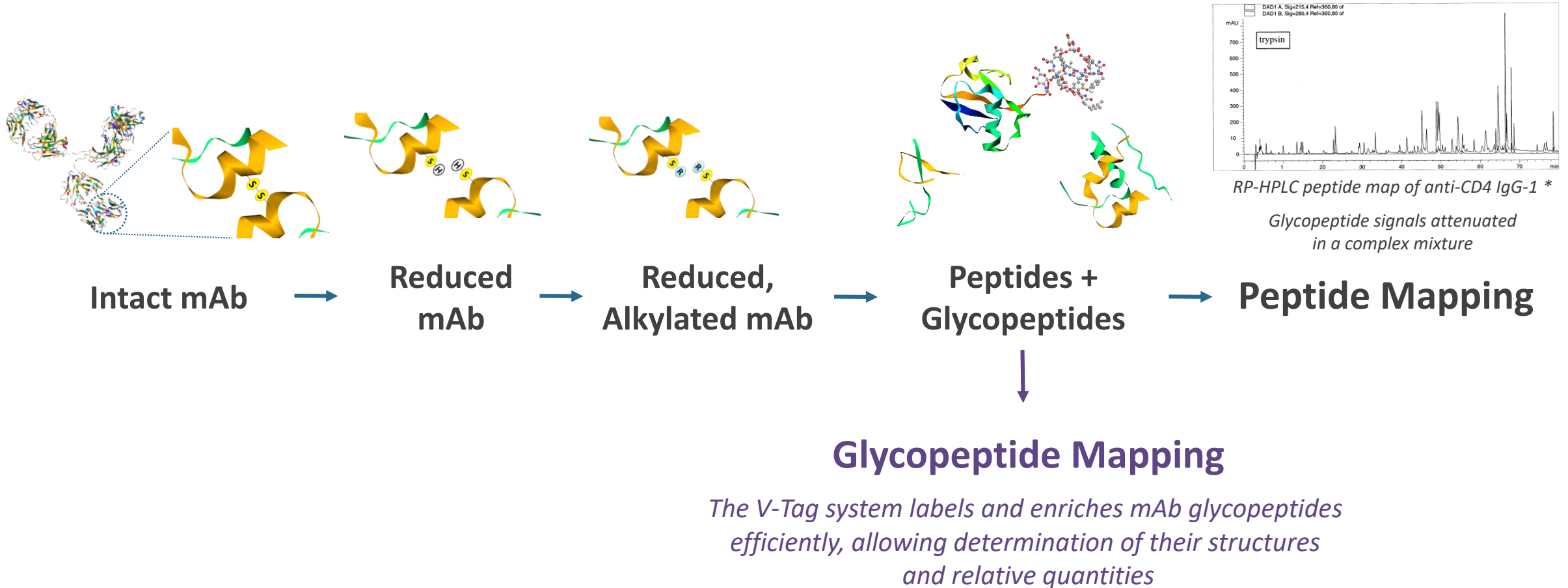
## 3. Glycans

*Using MS, UHPLC, CE*

- ✓ Provides **glycan identity and quantitation**
- X *Need to release glycans – extra work and expense. No glycosylation site information (for mAbs with Fab and Fc glycosylation)*

# Adding Glycoprofiling to the Traditional Peptide Mapping Workflow

Incorporating glycopeptide mapping can reduce the time and costs of glycoprofiling work in drug production monitoring



# Highlights of the V-Tag System

## Reliable mAb Glycoprofiling

*Allows glycan identification and quantitation using the orthogonal analyses of MALDI-MS and UHPLC. Provides data comparable to gold-standard glycoprofiling methods based on 2-AB or 2-AA*

## Minimal Sample Needed

*Excellent glycopeptide mapping using as little as 10 µg of mAb glycoprotein*

## Validated for GMP Labs

*Validated to ICH Q2(R1) standards and tested in glycoprofiling labs*



## Quick and Easy

*Labeling and enrichment is completed within 2 hours*

## Integrates Easily with Peptide Mapping Workflow

*Adds onto your existing peptide mapping workflow, without requiring extra steps for glycan release*

## Automatable for High-Throughput Studies

*The procedure is compatible with 96-well plate based assays, enabling high-throughput studies using a liquid handling robot*

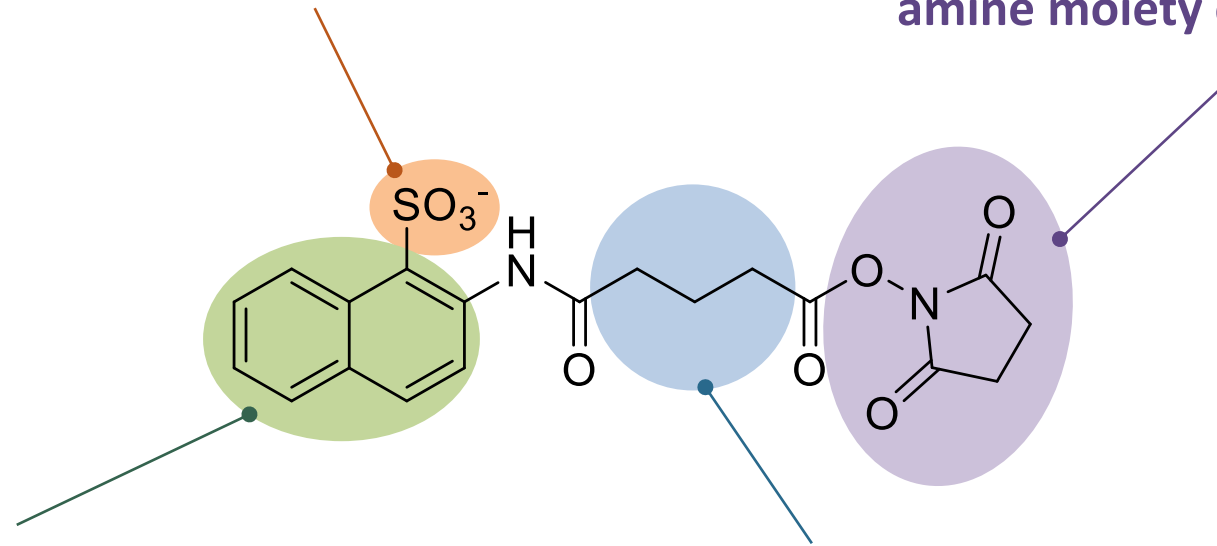
# Anatomy of V-Tag

The molecular aspects that make V-Tag work

**Sulphate** anion to improve analysis  
in negative ion mode on MALDI

**Amine reactive succinimidyl ester**  
to react with the N-terminus  
amine moiety of the peptide

**Fluorescent** group for  
detection in UHPLC  
 $\lambda_{\text{ex}} = 250\text{nm}$ ,  $\lambda_{\text{em}} = 360\text{nm}$



**Simple, non-reactive alkyl chain** to  
link the fluorescent moiety to the reactive  
succinimidyl ester

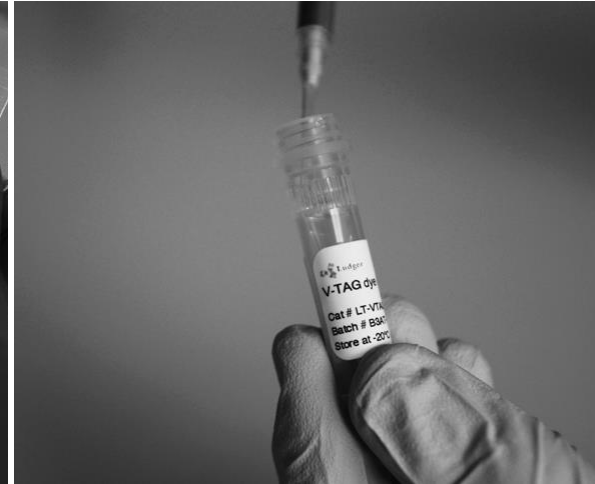
# Components of the LT-VTAG-24 Kit

## 1. Labeling



*Reaction buffer  
and solvent*

**PBS Buffer Tablet**  
LT-PBS-TAB-0.01M



*Amine reactive  
fluorescent labeling  
reagent*

**V-Tag Labeling Dye**  
LT-VTAG-01



*HILIC resin  
cartridge*

**LudgerClean A  
Cartridges (LC-A)**  
LC-A-24

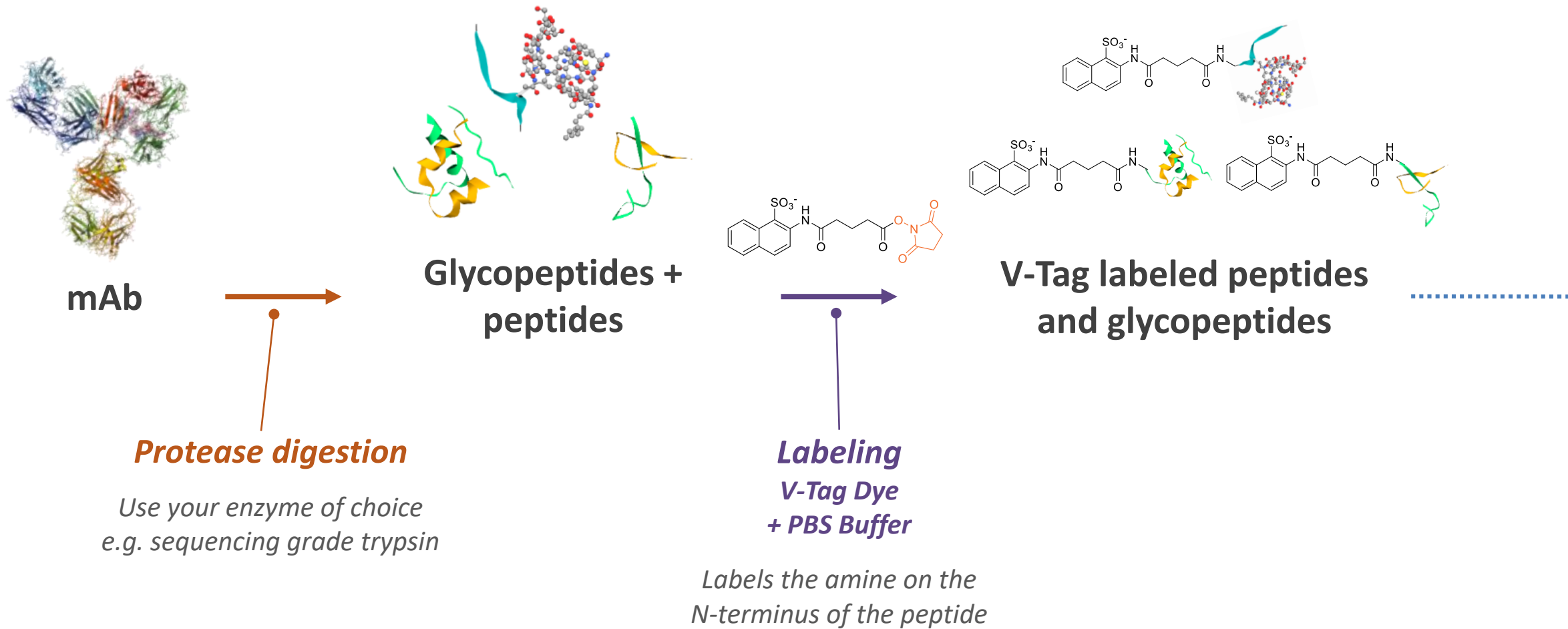
## 2. Enrichment



*Solvent  
involved in HILIC  
clean-up and enrichment*

**TFA 10% (aq.)**  
LC-TFA10PC-01

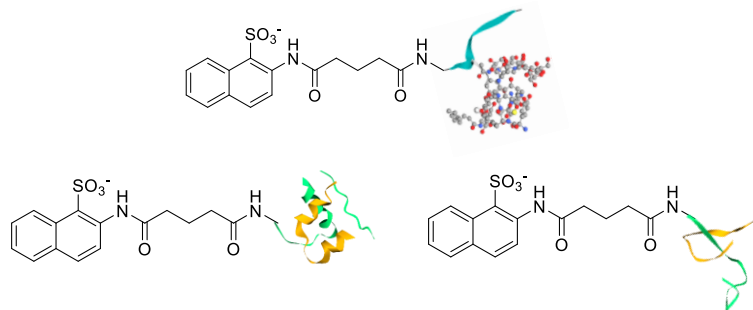
# Workflow for the V-Tag System: Stage 1 - Labeling





# Workflow for the V-Tag System: Stage 2 – Enrichment

Analysis by MALDI-MS and UHPLC

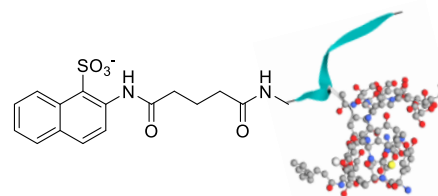


V-Tag labeled peptides  
and glycopeptides



**Enrichment**  
**LT-VTAG-24 Kit**  
**LC-A cartridges**  
**with TFA solutions**

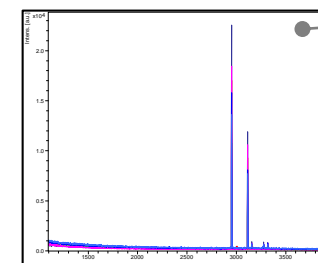
*Separation of mixture using HILIC  
cartridge. Conditions optimised for  
recovery of glycopeptides with  
glycosylation patterns preserved*



V-Tag labeled  
glycopeptides

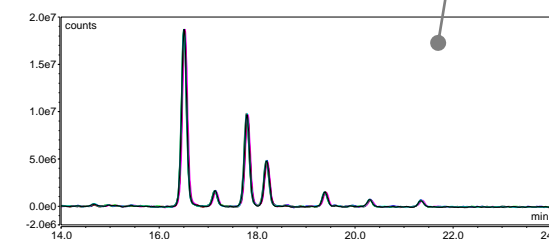


**Orthogonal**  
**Analyses**  
**MALDI-MS**  
**and UHPLC**



**MALDI-MS**  
**For glycan**  
**Identification**

**UHPLC**  
**Quantitation**



*IgG-1 V-Tag labeled glycopeptides*

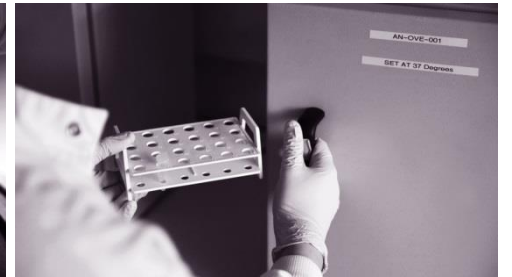
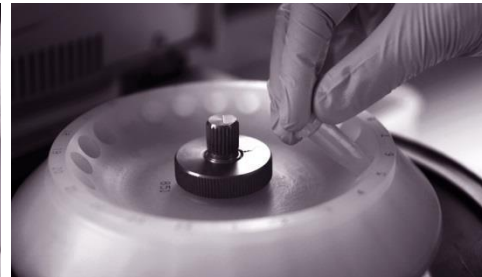
# V-Tag Workflow: Simple and Easy

Labeling

Buffering the mAb digest

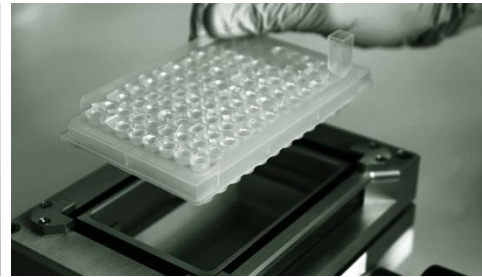


Label the digest

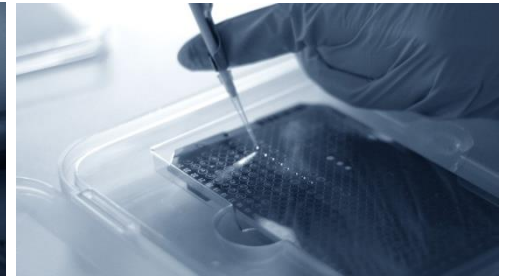
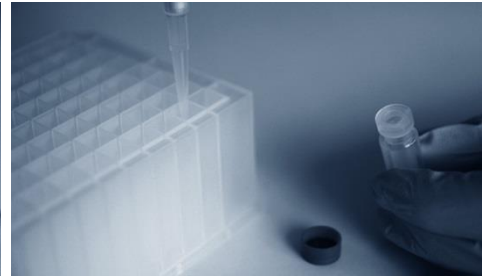
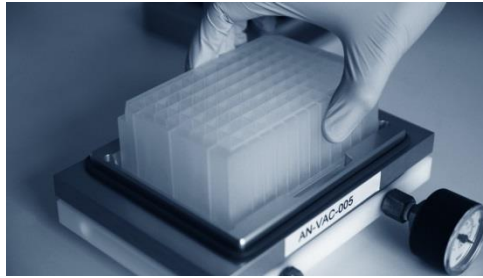
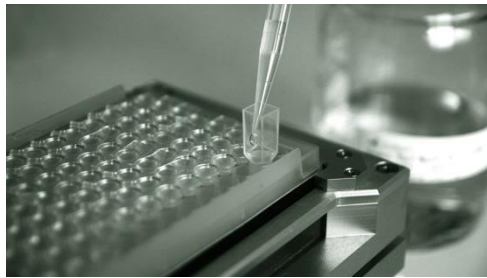


Enrichment

Remove peptides

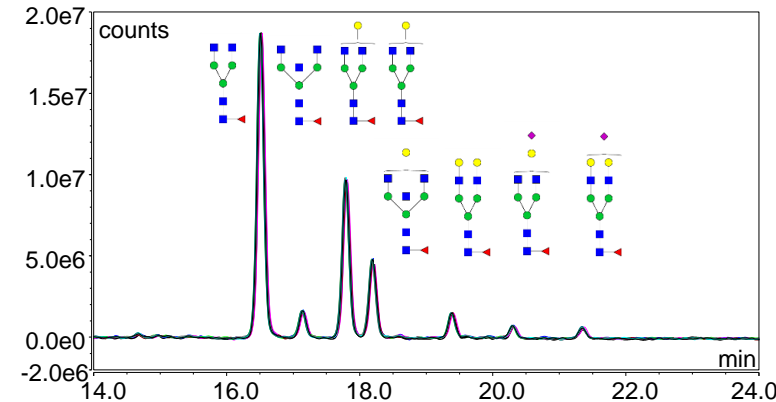
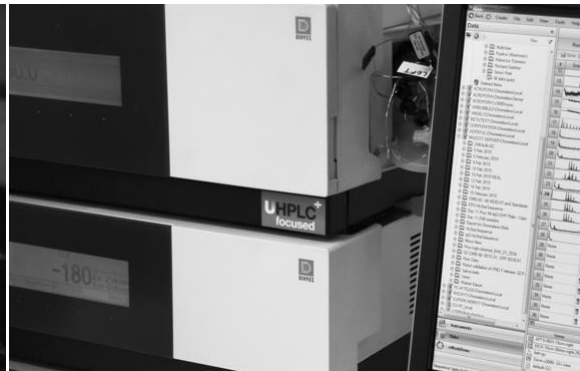


Elute glycopeptides



# Set-up of Orthogonal Glycoanalytical Platforms

Typical setup for analysis of V-Tag labeled glycopeptides by UHPLC and MALDI-MS



**Thermo Scientific  
Dionex U3000**  
30 minute gradient  
25  $\mu$ l injection

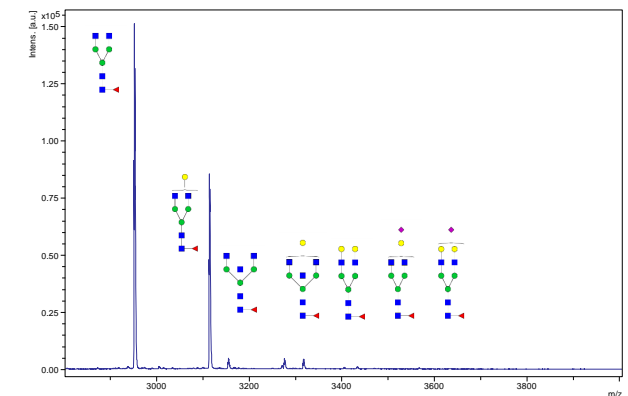
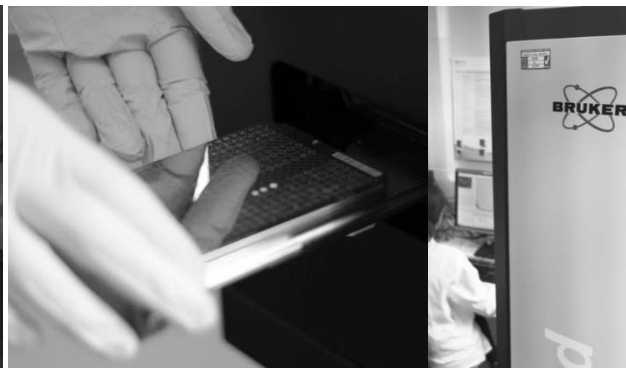
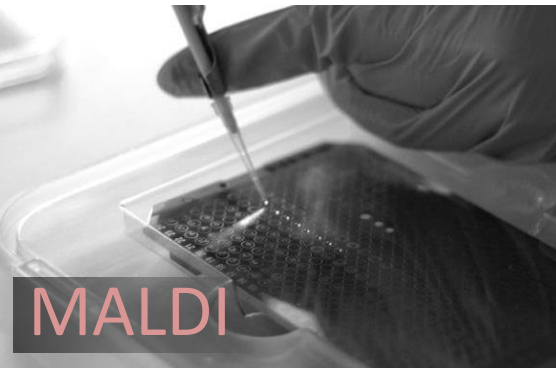


**Waters UHPLC Glycan  
BEH Amide Column (HILIC)**  
(150mm x 2.1mm)  
Temperature: 60  $^{\circ}$ C



**FLD  
Fluorescence Detector**  
 $\lambda_{ex} = 250 \text{ nm}$   
 $\lambda_{em} = 360 \text{ nm}$

**Typical data**  
V-Tag labeled glycopeptide map of IgG 1



**Spot Sample**  
Matrix: 2,5-dihydroxybenzoic  
acid (DHB)



**Load Plate**  
Bruker Autoflex  
MALDI-MS instrument

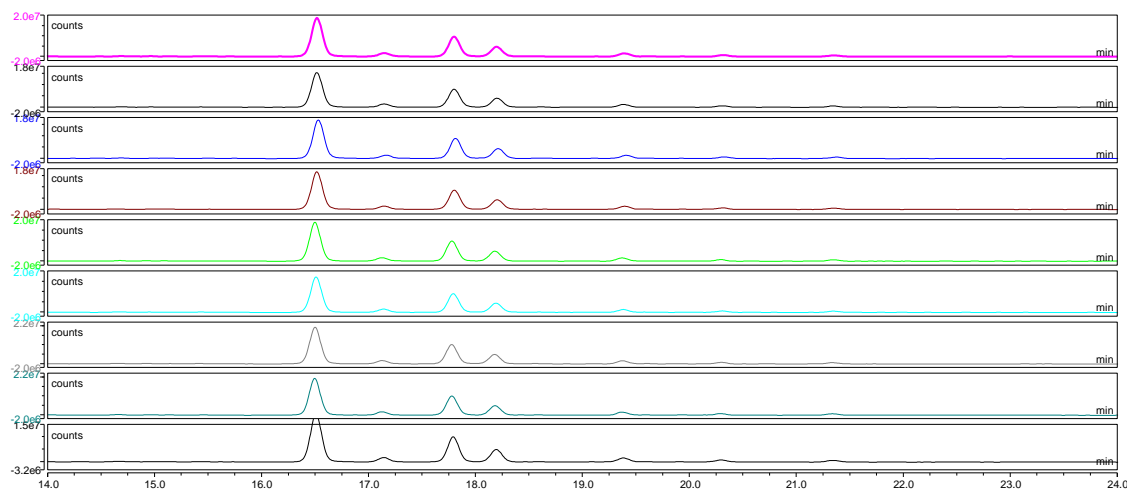


**Collect Data**  
Mode: reflectron  
negative ion

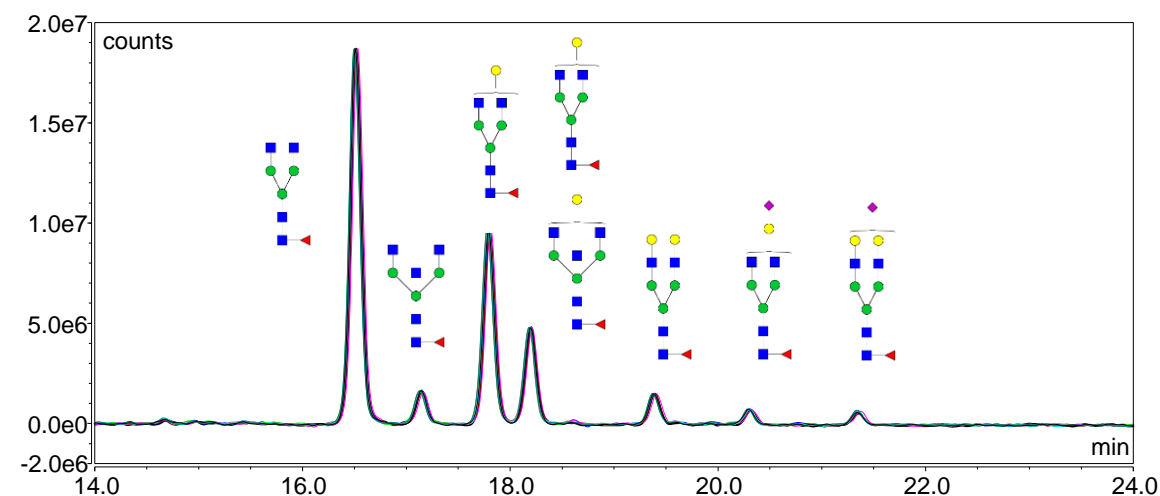
**Typical Data**  
V-Tag labeled glycopeptide map of IgG 1

# V-Tag System Has Been Validated to ICH Q2(R1) Level

V-Tag is reliable and robust and can be used for GMP level glycoprofiling of monoclonal antibodies



Stacked IgG 1 V-Tag labeled glycopeptide UHPLC profiles (9 replicates)



Overlaid IgG 1 V-Tag labeled glycopeptide UHPLC profiles (9 replicates)

Validation studies typically show repeatability with CVs for relative abundances < 4%

Peak Number		1	2	3	4	5	6	7
Glycopeptide		G0F (FA2)	G0FB (FA2B)	G1F (FA2G1)	G1F + G1FB (FA2G1 + FA2BG1)	G2F (FA2G2)	G1FS1 (FA2G1S1)	A1F (FA2G2S1)
Relative % Area	Av.	50.2	4.2	25.8	12.6	3.8	1.8	1.7
	Std. Dev.	0.35	0.14	0.18	0.26	0.10	0.02	0.04
	CV	0.70	3.20	0.69	2.05	2.54	0.92	2.40

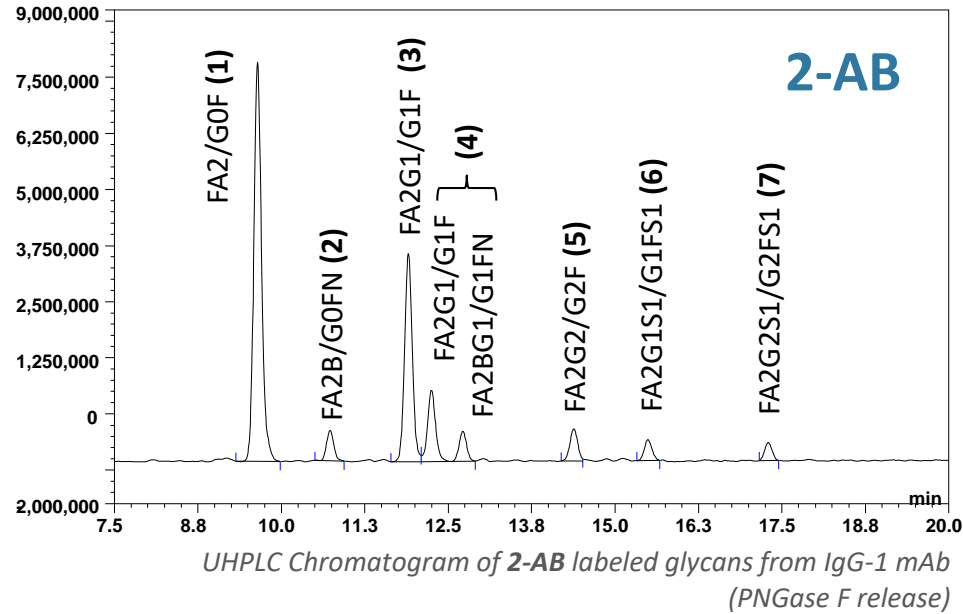
Average relative % area, SD and CVs for V-Tag labeled IgG 1 glycopeptides



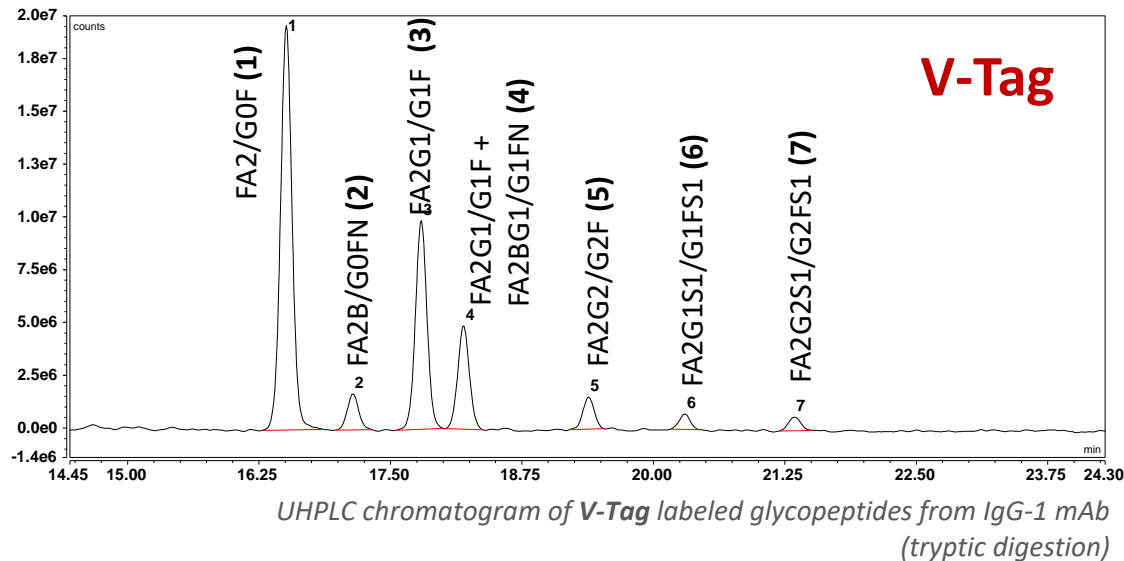
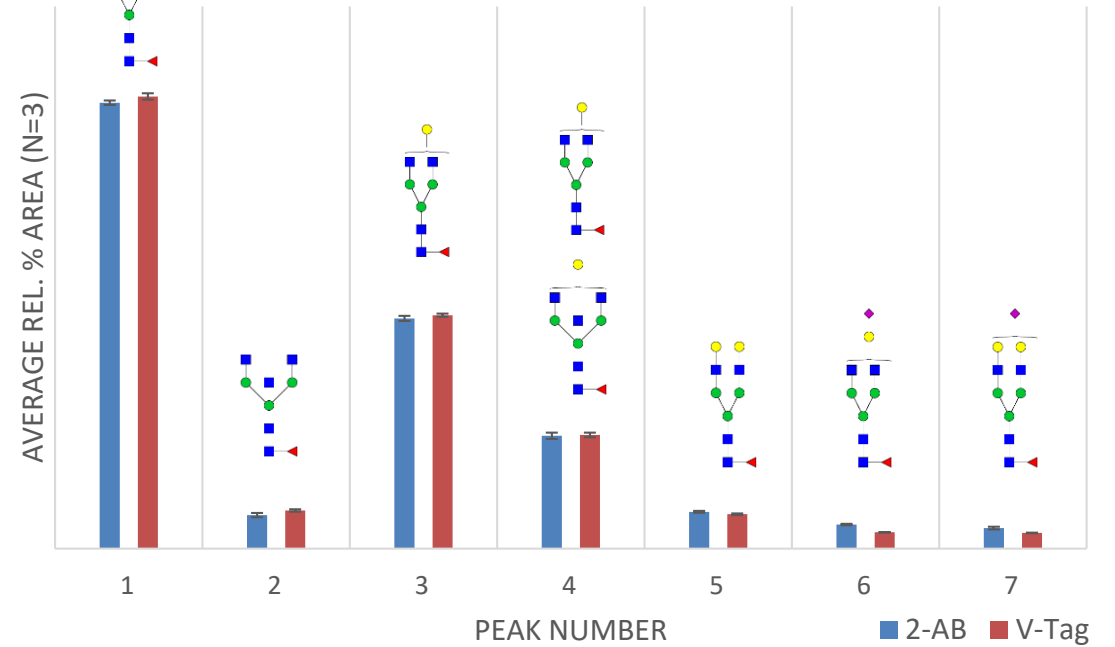
Studies using  
V-Tag at Ludger

# Comparability of 2-AB and V-Tag

V-Tag labeling of glycopeptides gives comparable results to the gold standard 2-AB labeling of glycans

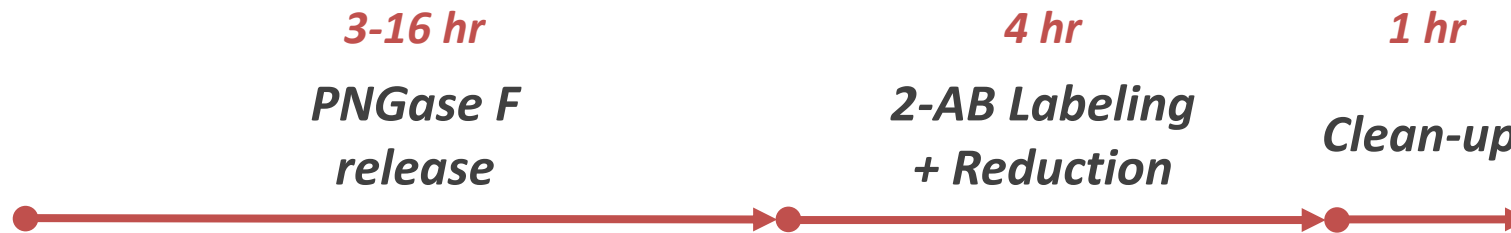


Comparable in quantitation and in reliability, using a Waters UHPLC Glycan BEH Amide Column (HILIC)

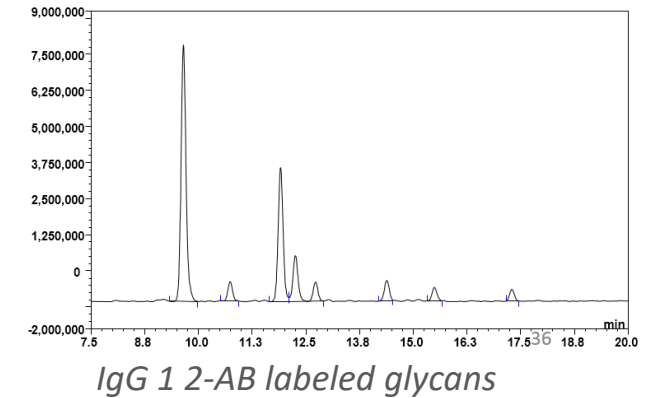


# The V-Tag protocol is much shorter than 2-AB

## 2-AB labeling of N-Glycans



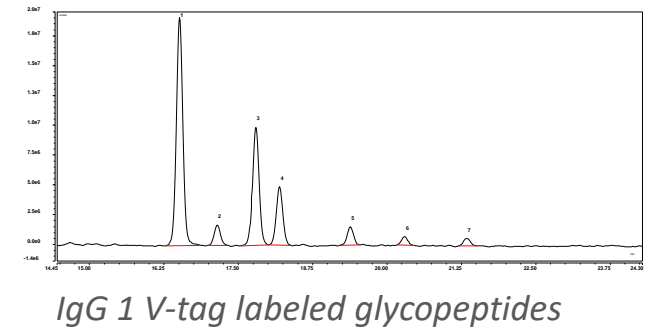
Total: 8 -21 hrs



## V-Tag labeling of glycopeptides



Total: 3 hrs

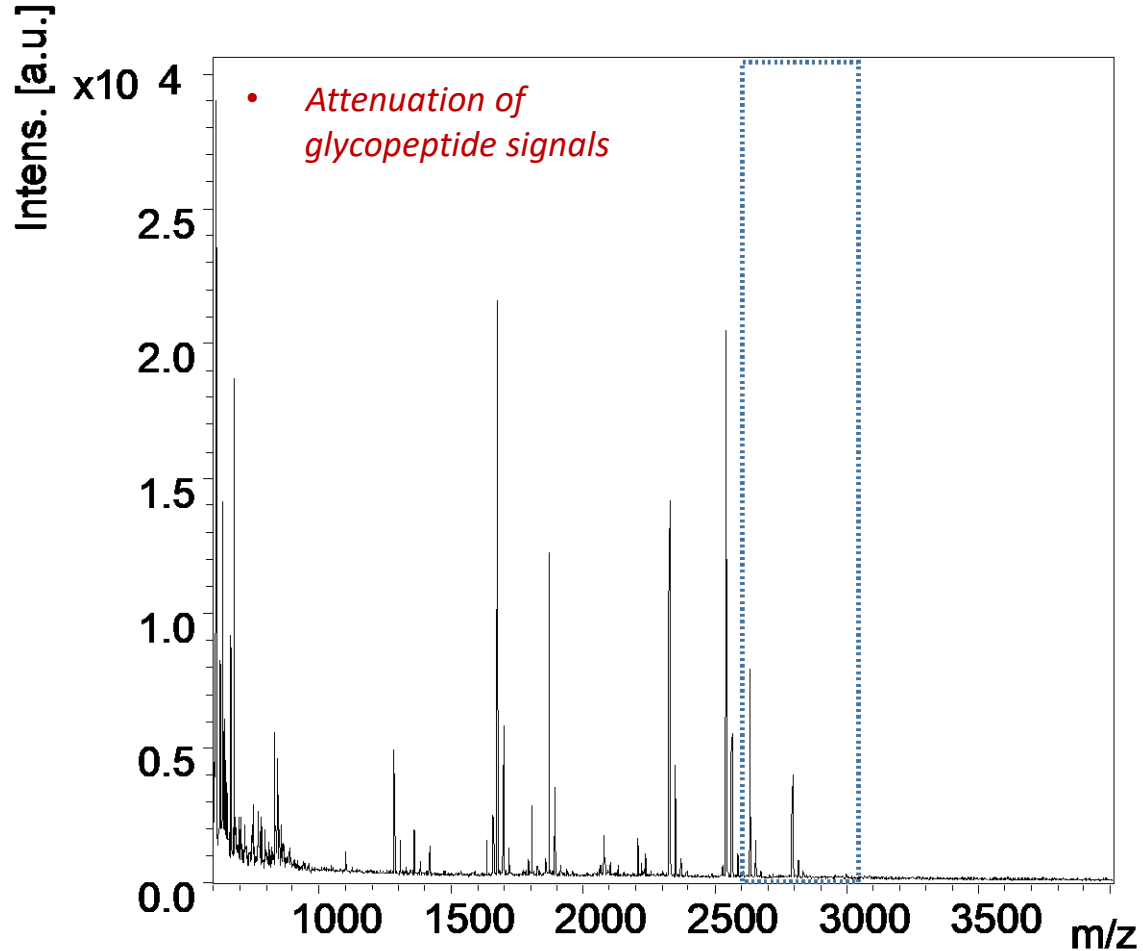


# V-Tag Greatly Enhances MALDI-MS Analysis of Glycopeptides

The signal for underderivatised glycopeptides is suppressed in MALDI-MS but is enhanced after V-tag labeling and enrichment

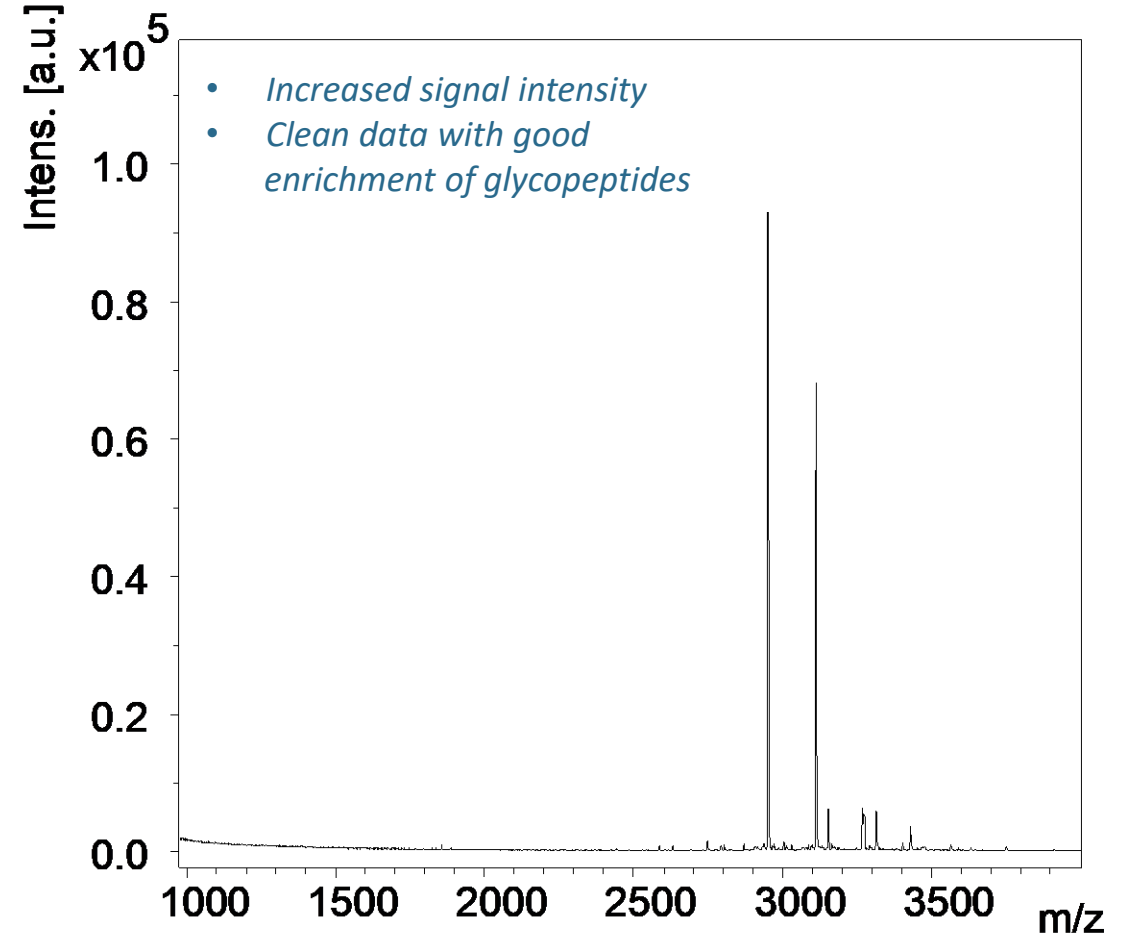
## Before V-Tag labeling and enrichment

*IgG 1 tryptic peptides and glycopeptides*



## After V-Tag labeling and enrichment

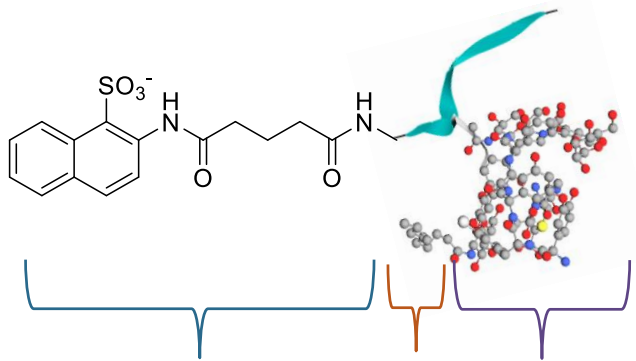
*IgG 1 tryptic glycopeptides*





# V-Tag Allows Identification of Glycopeptides using MALDI-MS

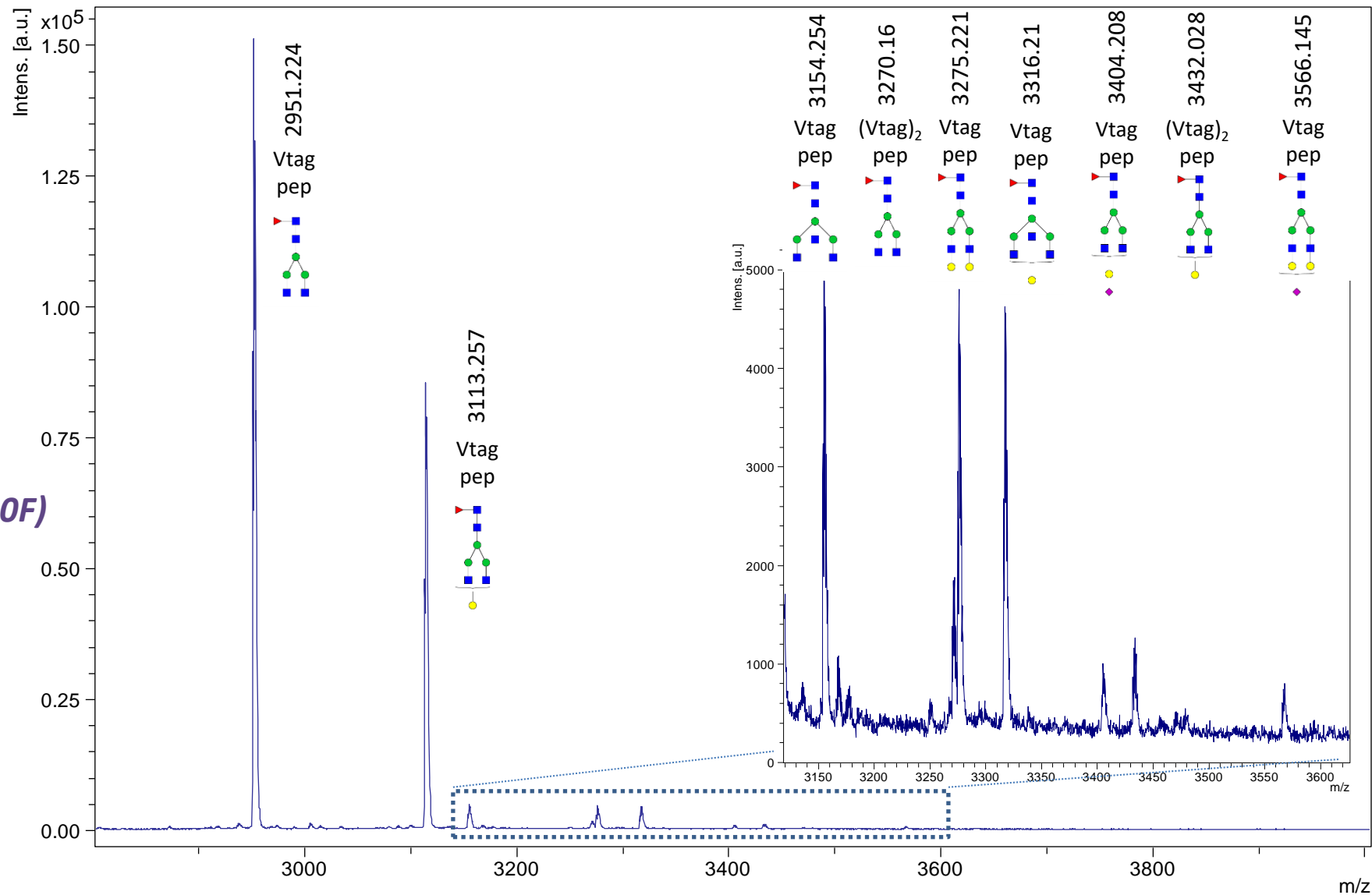
Example of V-Tag labeled IgG-1 mAb glycopeptide analysis by negative mode MALDI-MS



$M = \text{V-Tag} + \text{EEQYNSTYR} + \text{Glycan}$

$M = 319.33 + 1189.52 + 1444.53 \text{ (GOF)}$

$M = 2952.11$



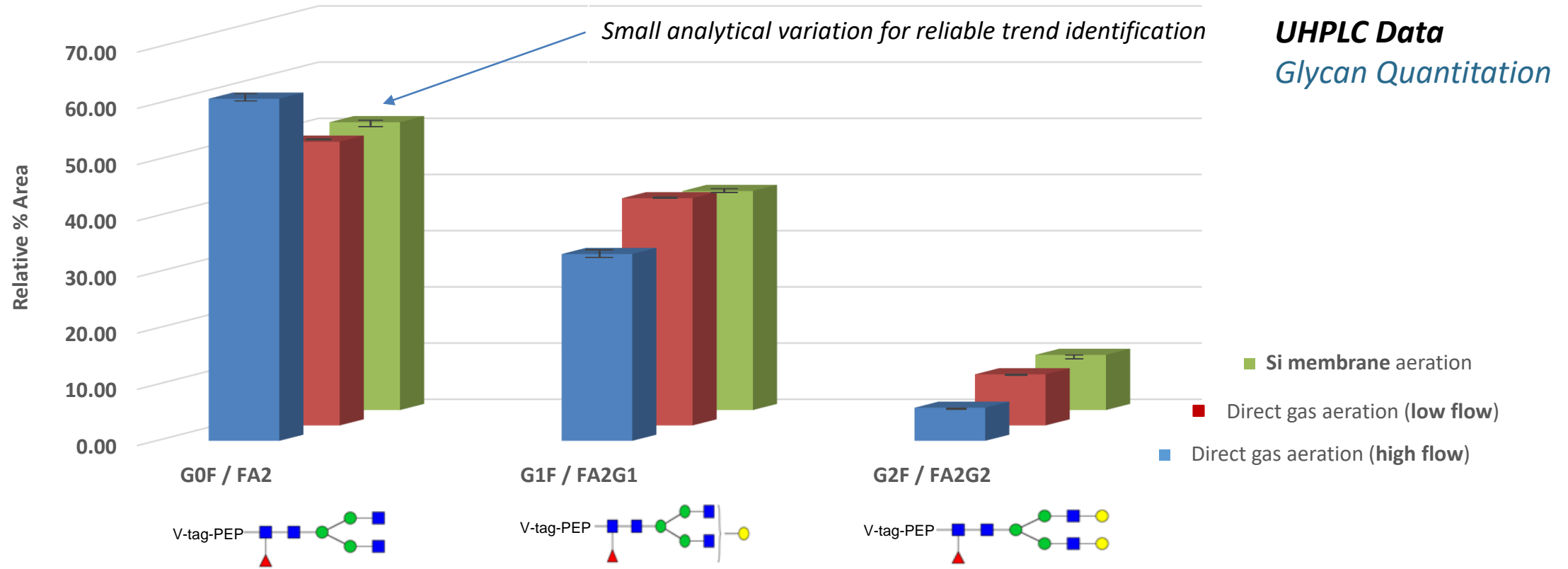
MALDI-MS ion signals for V-Tag labeled IgG-1 glycopeptides (negative ion mode; Bruker Autoflex)

Incorporating  
V-Tag into your  
drug programme



# QbD Study: The impact of cell culture conditions on glycoform patterns

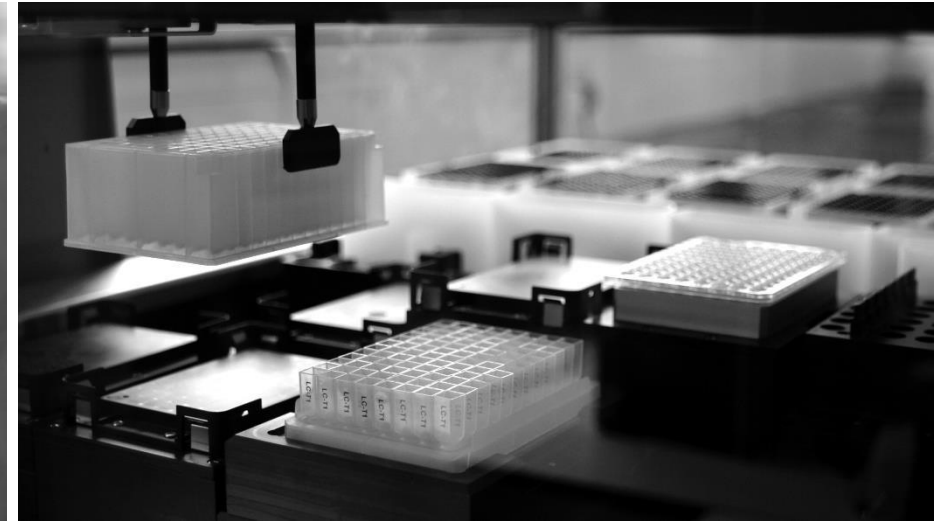
Variations are reliably detected with V-tag system



- Chinese hamster ovary (CHO) cell line GS-CY01 expressing a mAb was grown in bio-reactors using *different aeration conditions*
- Fc galactosylation patterns analysed (i.e. the ratios of the G0F, G1F and G2F glycans) for different aeration conditions.
- Increasing the levels of terminal galactose are known to positively correlate with complement dependent cytotoxicity (CDC) activity
  - The cells grown under silicon membrane aeration showed the highest degree of Fc galactosylation (higher abundance of G2F)

# Automated High Throughput Studies using V-Tag

Adapted to 96-well plate system to use with a liquid handling robot



..... Morning .....

..... Afternoon or Overnight .....



*The workflow can be completed in **1 day** making this technology a good candidate for high throughput analysis of mAbs*

# Next Steps...

If you have a question



CLICK  
to contact  
Jenifer

**Dr Jenifer Hendel**

*Senior Scientist*

*jenifer.hendel@ludger.com*

Request a quotation



CLICK  
to contact  
Sales

**Sales Team**

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*Orders: sales@ludger.com*